

## Letter

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### Comments on: *Mutations in Mitochondrial Control Region DNA in Gastric Tumours of Japanese Patients, Tamura, et al. Eur J Cancer 1999, 35, 316-319*

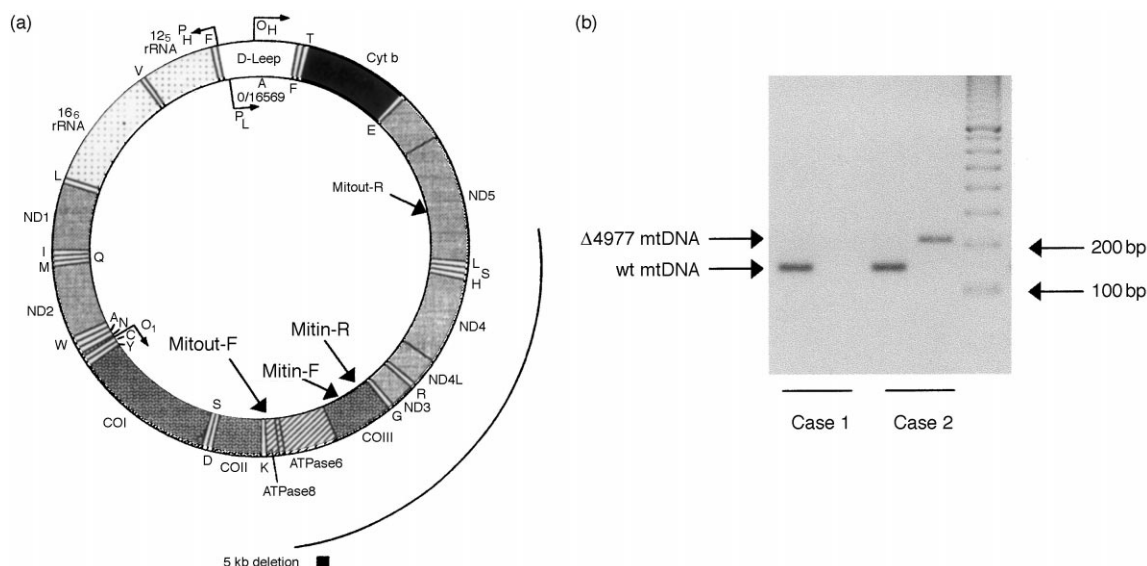
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IN A recent issue of the *European Journal of Cancer*, Tamura and colleagues [1] reported mutations in mitochondrial control region DNA in 2/45 gastric tumours (4.4%). The two mutations were detected in the hypervariable regions HV1 and HV2; a G to A transition at nucleotide position 16129

and a 1 bp A deletion at nucleotide position 248, respectively. Despite the high percentage of replicative error negative (RER-) tumours (only 8/45 tumours exhibited an RER+ phenotype), the two cases with mtDNA mutations were RER- carcinomas. Tamura and colleagues [1] suggested that the observation of carcinomas with mtDNA instability that do not exhibit nuclear microsatellite instability might be explained by alterations of a mismatch repair system that would act specifically in mitochondrial genome integrity, namely a MSH1-like enzyme [2].

The results we have recently obtained partly confirm and partly diverge from the results reported by Tamura and colleagues [1]. We searched for the mitochondrial common deletion ( $\Delta$ 4977 mtDNA) in a series of 32 primary gastric carcinomas. 10/32 primary gastric carcinomas (31.3%) had microsatellite instability (RER+ phenotype) evaluated according to the methods previously described [3]. The detection of  $\Delta$ 4977 mtDNA was performed on DNA extracted from microdissected frozen tissues as previously described [4], using two sets of primers: Mitout-F -5'-cccaactaaatactaccgtatgg-3' and Mitout-R -5'-ggctcaggc-gttgtgtatgat-3' (outside the  $\Delta$ 4977 mtDNA) and Mitin-F -5'-ctgagcctttaccactccag-3' and Mitin-R -5'-ggtgattgata-ctcctgatgcg-3' (within the  $\Delta$ 4977 mtDNA). In the wild type mtDNA (wt mtDNA) only the Mitin primer set gives a PCR product with 142 bp. In cases with the  $\Delta$ 4977 mtDNA, Mitin primers amplify a 142 bp target sequence and Mitout primers an aberrant PCR product with 214 bp (Figure 1). We detected  $\Delta$ 4977 mtDNA in 17/32 primary sporadic gastric carcinomas (53.1%) (Table 1). The high frequency of large mtDNA deletions found in our series in comparison with the low frequency of mtDNA point mutations in the series of Tamura and colleagues [1], suggests that gastric cancer is more prone to have gross genetic alterations of



**Figure 1. (a) Human mtDNA map showing the location of mitochondrial genes. Arrows represent the localisation of the primers (Mitin and Mitout) used for amplification. The out line represents the  $\Delta$ 4977 mtDNA. (b) Analysis of mtDNA PCR products. Case 1 showing only Mitin amplification is negative for  $\Delta$ 4977 mtDNA and Case 2 showing both Mitin and Mitout amplification is positive for  $\Delta$ 4977 mtDNA.**

Table 1. Relationship of RER phenotype and  $\Delta 4977$  mtDNA in 32 gastric carcinomas

RER + phenotype	Del Mit + (n = 17)	Del Mit- (n = 15)	Total (n = 32)	P value*
Positive	1	9	10	0.0010
Negative	16	6	22	

\*P value was obtained using the Chi-square test with Yates correction.

mtDNA than to exhibit signs of fine genetic instability. This assumption is supported by the findings of Burgart and colleagues [5], who detected a 50bp mtDNA deletion in 4/32 gastric carcinomas (12.5%) of the cardia. The same does not hold true in colorectal carcinomas where somatic point mutations of mtDNA appear to be much more prevalent than mtDNA deletions [6].

In keeping with the results of Tamura and colleagues [1] who found mtDNA mutations only in two RER- tumours, almost every carcinoma of our series displaying  $\Delta 4977$  mtDNA were also RER- (16/17 tumours) (94.1%) ( $P = 0.001$ ) (Table 1).

The inverse association found by Tamura and colleagues [1] and ourselves, between RER+ phenotype and mitochondrial alterations (somatic point mutations and  $\Delta 4977$  mtDNA), remains to be clarified. We think it cannot be ascribed to a different DNA mismatch repair gene defect,

namely MSH1, as it was suggested by Tamura and colleagues [1] because if that were the case, one would expect the occurrence of mtDNA alterations regardless of genetic nuclear instability. Alternatively, we think it is tempting to hypothesise that such an inverse relationship may reflect a putative lethal effect caused on the neoplastic cells by the association of mtDNA deletions and DNA repair deficiency, but this possibility remains to be proven.

1. Tamura G, Nishizuka S, Maesawa C, *et al.* Mutations in mitochondrial control region DNA in gastric tumours of Japanese patients. *Eur J Cancer* 1999, **35**, 316–319.
2. Reenan RA, Kolodner RD. Isolation and characterization of two *saccharomyces cerevisiae* genes encoding homologs of the bacterial HexA and MutS mismatch repair proteins. *Genetics* 1992, **132**, 963–973.
3. Oliveira C, Seruca R, Seixas M, Sobrinho-Simões M. The clinicopathological features of gastric carcinomas with microsatellite instability may be mediated by mutations of different “target genes”: a study of the TGFbeta RII, IGFIR, and BAX genes. *Am J Pathol* 1998, **153**, 1211–1219.
4. Máximo V, Soares P, Rocha AS, Sobrinho-Simões M. The common deletion of mitochondrial DNA is found in goiters and thyroid tumors with and without oxyphil cell change. *Ultrastruct Pathol* 1998, **22**, 271–273.
5. Burgart LJ, Zheng J, Shu Q, Strickler JG, Shibata D. Somatic mitochondrial mutation in gastric cancer. *Am J Pathol* 1995, **147**, 1105–1111.
6. Polyak K, Li Y, Zhu H, *et al.* Somatic mutations of the mitochondrial genome in human colorectal tumours. *Nat Genet* 1998, **20**, 291–293.